

PLANT CELL WALL MONOCLONAL ANTIBODIES



>50 rat antibody hybridoma cell lines

Prof. Paul Knox, emeritus professor

University of Leeds, UK

email: j.p.knox@leeds.ac.uk

www.plantcellwalls.leeds.ac.uk

Notes on the selection and use of our MAbs

The LM- and other series of monoclonal antibodies (MAbs) developed in Leeds is a large resource of probes for the analysis of plant cell wall carbohydrates. If you require a specific antibody/epitope then that can be straightforward. If you wish to study and explore wider aspects of cell walls using a panel of antibodies that can detect a range of glycans then we have series of **recommended MAbs** for polysaccharide classes.

For pectic HG we recommend the combined use of **LM19** (unesterified HG) and **LM20** for high methylester HG. **JIM7** is also always additionally recommended for any analysis as it binds widely to pectin.

For detection of xyloglucan we recommend the high affinity **LM25** and for heteromannan **LM21**.

There is a range of probes for heteroxylan – the core two to use would be **LM11** that binds to 1,4-xylosyl residues and **LM28** to glucuronosyl residues. The often different binding patterns of **LM10** and **LM11** is now known to arise from the specific recognition by **LM10** of the non-reducing end (NRE) of xylans.

It can be hard to predict which AGP MAb to select as these glycan epitopes vary between tissues, organs and species. **JIM13** and **LM2** are a good place to start as they usually detect something in a section or an extract. Do not forget that a single AGP glycan epitope is unlikely to detect all AGPs in an organ. For starting with extensins we suggest the use of **LM1** and **JIM20**.

Pectic polysaccharides

Homogalacturonan (HG) / related

LM19 partially Me-HG / no ester (34)

LM20 partially Me-HG (34)

JIM7 partially Me-HG (6,22,26)

LM7 partially Me-HG / non-blockwise, (24,26)

JIM5 partially Me-HG / no ester (1,6,22,26)

LM18 partially Me-HG / no ester (34)

PAM1 blockwise de-esterified HG (17,22,23,30)

LM8 xylogalacturonan (27)

Rhamnogalacturonan-I

Galactan +

LM5 NRE (1→4)-β-D-galactan (15,18-20,42)

LM26 branched (1,6-Gal) (1→4)-β-D-galactan (45)

Arabinan +

LM6⁺ (1→5)-α-L-arabinan (16,18-20,23,31,34)

LM6-M (1→5)-α-L-arabinan (46)

LM13 linearised (1→5)-α-L-arabinan (33,35)

Other

LM16 processed arabinan/put. galactan stub (35)

LM9 feruloylated (1→4)-β-D-galactan (28)

LM12[‡] ferulic acid, feruloylated pectin (38)

⁺May also bind to AGPs

[‡]Can also bind to feruloylated heteroxylan

Non-cellulosic, non-pectic polysaccharides

Xyloglucan

LM15 XXXG motif of xyloglucan (32,44)

LM24 galactosylated xyloglucan (38)
LM25 XXXG/galactosylated xyloglucan (38)
Heteromannan
LM21 heteromannan (36)
LM22 heteromannan (36)
Heteroxylan

LM10 NRE (1→4)-β-D-xylan (29,44)
LM11 (1→4)-β-D-xylan / arabinoxylan (29)
LM28 glucuronoxylan (40)
LM12[‡] ferulic acid, feruloylated xylan (38)
LM27 unknown epitope assoc. grass xylan (40)
[‡]Can also bind to feruloylated pectin

Plant cell wall proteoglycans/glycoproteins

Arabinogalactan-protein (AGP) glycan

LM2 β-linked-GlcA in AGP glycan (13,14)
LM14 GlcA in AGP glycan (33,38)
LM30 AGP glycan (43)
JIM4 AGP glycan (3,5,13)
JIM13 AGP glycan (7,13)
JIM14 AGP glycan (7,13,44)
JIM15 AGP glycan (7,13)
JIM16 AGP glycan (7,13,44)
MAC207 AGP glycan (2,3,13)

Extensin

LM1 extensin (11)
JIM11 extensin (8)
JIM12 extensin (8)
JIM19 extensin (8,9,10)
JIM20 extensin (8,9)

NRE = epitope at non-reducing end of glycan

Other cell wall related MAbs

LM23 non-acetylated xylosyl in xylogalacturonan, xylan, fucoidan preps (37,38, 39)
LM4 pea amine oxidase, cell walls (21,25)
JIM18 glyco-phospholipid, membranes (9,12)
JIM1 β-linked-galactosyl, plasma membrane (4)

Brown algal cell wall polysaccharides

Fucoidan

BAM1 un-sulfated epitope present in sulfated fucan/fucoidan preparations (39)
BAM2 sulfated epitope present in sulfated fucan/fucoidan preparations (39)
BAM3 possibly sulfated epitope present in sulfated fucan/fucoidan preparations (39)
BAM4 sulfated epitope present in sulfated fucan/fucoidan preparations (39)

Alginate

BAM6 manuronate-rich epitope (41)
BAM7 manuronate-guluronate (41)
BAM8 manuronate-guluronate (41)
BAM9 manuronate-guluronate (41)
BAM10 manuronate-guluronate epitope resistant to alginate lyase (41)
BAM11 ~7 guluronate residues (41)

If you have specific questions about our MAb specificities or use
please enquire at j.p.knox@leeds.ac.uk

References

1. VandenBosch *et al.* (1989) **EMBO Journal** 8, 335-342
2. Pennell. *et al.* (1989) **J. Cell Biology** 108, 1967-1977
3. Knox *et al.* (1989) **Development** 106, 47-56
4. Knox & Roberts (1989) **Protoplasma** 152, 123-129
5. Stacey *et al.* (1990) **Planta** 180, 285-292
6. Knox *et al.* (1990) **Planta** 181, 512-521
7. Knox, *et al.* (1991) **Plant Journal** 1, 317-326
8. Smallwood *et al.* (1994) **Plant Journal** 5, 237-246
9. Knox *et al.* (1995) **Planta** 196, 266-270
10. Wang *et al.* (1995) **Planta** 196, 271-276
11. Smallwood *et al.* (1995) **Planta** 196, 510-522
12. Perotto *et al.* (1995) **Mol. Plant-Microbe Int.** 8, 560-568
13. Yates *et al.* (1996) **Glycobiology** 6, 131-139
14. Smallwood *et al.* (1996) **Planta** 198, 452-459
15. Jones *et al.* (1997) **Plant Physiology** 113, 1405-1412
16. Willats *et al.* (1998) **Carbohydr. Research** 308, 149-152
17. Willats *et al.* (1999) **Plant Journal** 18, 57-65
18. Willats *et al.* (1999) **Plant Journal** 20, 619-628
19. McCartney *et al.* (2000) **Plant Journal** 22, 105-113
20. Orfila & Knox (2000) **Plant Physiology** 122, 775-781
21. Wisniewski *et al.* (2000) **Mol. Plant-Microbe Int.** 13, 413
22. Willats *et al.* (2000) **Carbohydr. Res.** 327, 309-320
23. Orfila *et al.* (2001) **Plant Physiology** 126, 210-221
24. Willats *et al.* (2001) **J. Biol. Chem.** 276, 19404-19413
25. Laurenzi *et al.* (2001) **Planta** 214, 37-45
26. Clausen *et al.* (2003) **Carbohydr. Res.** 338, 1797-1800
27. Willats *et al.* (2004) **Planta** 218, 673-681
28. Clausen *et al.* (2004) **Planta** 219, 1036-1041
29. McCartney *et al.* (2005) **J. Histochem Cytochem** 53, 543
30. Manfield *et al.* (2005) **Plant Science** 169, 1090-1095
31. Lee *et al.* (2005) **Plant Cell** 17, 3051-3065
32. Marcus *et al.* (2008) **BMC Plant Biology** 8, 60
33. Moller *et al.* (2008) **Glycoconjugate J.** 25, 37-48
34. Verhertbruggen *et al.* (2009) **Carbohydr. Res.** 344, 1858
35. Verhertbruggen *et al.* (2009) **Plant Journal** 59, 413-425
36. Marcus *et al.* (2010) **Plant Journal** 64, 191-203
37. Manabe *et al.* (2011) **Plant Physiology** 155, 1068-1078
38. Pedersen *et al.* (2012) **J. Biol. Chem.** 287, 39429-39438
39. Torode *et al.* (2015) **PlosONE** 10, e0118366
40. Cornuault *et al.* (2015) **Planta** 242,1321-1334
41. Torode *et al.* (2016) **J. Exp Botany** 67, 6089-6100
42. Andersen *et al.* (2016) **Carbohydrate Research** 436, 36-40
43. Wilkinson *et al.* (2017) **J. Cereal Science** 74, 155-164
44. Ruprecht *et al.* (2017) **Plant Physiology** 175, 1094-1104
45. Torode *et al.* (2018) **Plant Physiology** 176, 1547-1558
46. Cornuault *et al.* (2018) **BioRxiv** /10.1101/161604

MAB selections available through

Kerafast

<https://www.kerafast.com/cat/799/paul-knox-phd>

Biosupplies Australia

<https://www.biosupplies.com.au/products/>

Absolute Antibody

Recombinant forms being made available: currently LM5, LM26, LM19 & LM20

<https://absoluteantibody.com/>

Brown algal BAM-series of MABs available from SeaProbes

<http://www.sb-roscoff.fr/en/seaprobes>

Summer 2026